

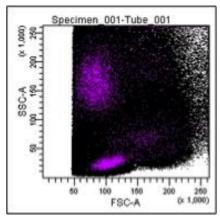


# DR5.APPNOTE: FLOW CYTOMETRY 002 060323

# **1. NUCLEATED CELL GATING**

### BACKGROUND

Normal blood and bone marrow samples contain a mixture of nucleated cells (leukocytes - including lymphocytes, monocytes, neutrophils - and various progenitor cells) and enucleated cells (mature erythrocytes and platelets). Typically, there are around 40 platelets and 400 erythrocytes for every nucleated cell. These can interfere with analyses of the nucleated cells, especially in flow cytometry where they complicate and slow phenotypic analysis. The most common solution is to osmotically shock the enucleated cells with  $NH_4CI - known$  as RBC lysis. After this, the nucleated cells are pelleted by centrifugation, washed, counted and resuspended for use.



### WHAT IS THE PROBLEM?

There are many potential risks from RBC lysis: additional time required; release of debris into the sample that can aggregate with leukocytes; inconsistent results; possible lysis of erythroid precursors; non-specific cell losses during washing procedures. The relative importance of these may vary but obviously sample loss would be of concern with mouse tail vein samples or in the detection of extremely rare cells such as circulating tumor cells, for example.

### HOW DOES DRAQ5<sup>™</sup> HELP?

The presence of genomic DNA is a simple way to differentiate between nucleated and enucleated cells. DRAQ5<sup>™</sup> is a live-cell permeant dsDNA-specific probe that efficiently and stably labels nucleated cells. It is added to diluted whole blood or bone marrow, mixed and briefly incubated. DRAQ5<sup>™</sup> fluoresces in the far-red when excited by blue or red laser on standard flow cytometer. The signal is detected in any channel above 675 nm, preferably including the peak emission at 697 nm. This signal is then used to select exclusively or "gate" the nucleated cells without the complexity and risk associated with RBC lysis. The nucleated cell population may include rare progenitors like reticulocytes, or circulating dysplastic cells (e.g. in MDS, PNH), "occult" tumor cells or endothelial cells not present in healthy blood or marrow. These are better preserved by avoiding RBC lysis.

Additionally, the DRAQ5<sup>™</sup> signal can identify cell doublets (plotting peak area versus peak width) and DNA content of each cell (as described below). The far-red fluorescence of DRAQ5<sup>™</sup> means that it can be combined with most visible range chromophores with limited or no spectral overlap.



For a full price list and further information see www.biostatus.com or contact us at: BioStatus Limited 56a Charnwood Road, Shepshed, Leicestershire LE12 9NP United Kingdom T +44 1509 558 163 | F +44 1509 651 061 | E enquiry@biostatus.com | W www.biostatus.com





# 2. DNA CONTENT (OR CELL CYCLE) ANALYSIS

## BACKGROUND

The cell cycle describes a series of events that occur when a cell divides and replicates into two daughter cells. Cell cycle regulation depends upon processes that are pivotal to cell survival. During the cell cycle the quantity of DNA increases from 2N (G1 phase) to 4N (G2 phase) via the S (synthesis) phase. Perturbation of the cell cycle can lead, for example, to cell cycle arrest or uncontrolled cell division, as in cancer, while cells with DNA below 2N are typically in apoptosis. The quantity of DNA in each nucleated cell in a population can be measured by labeling the nuclear DNA with a fluorescing DNA dye, most commonly propidium iodide (PI). The signals are plotted to create a DNA profile for the population giving information on the proliferative status, usually alongside phenotyping.

### WHAT IS THE PROBLEM?

Using PI to measure DNA content requires that nucleated cells are separated from a blood or bone marrow sample by ficoll density centrifugation. Thereafter, the isolated cells (or cultured cells) are then permeabilized (to allow dye entry) and treated with RNase (PI non-specifically binds to RNA). Spectrally PI is excited by the blue laser and detected around 610 nm, overlapping with R-PE a very bright and commonly used antibody conjugated dye. Alternatively, one can use the live-cell permeant DNA probe Hoechst 33342 that is excited by UV light. However, UV sources are less common on modern flow cytometers, replaced by violet sources driven by the emergence of new classes of violet–excited antibody labels.

### HOW DOES DRAQ5<sup>™</sup> HELP?

As a live-cell permeant DNA probe DRAQ5<sup>™</sup> can be applied directly to complex samples such as blood or bone marrow (as described above), as <u>the last step prior to analysis and without</u> <u>washing</u>. No complex processing, such as RBC lysis or ficoll separation and permeabilization, is needed. Its high dsDNA specificity avoids the need for RNase treatment. It is possible to get DNA profiles with the minimum disturbance to the sample, especially important for precious, small samples or where there is a risk of uncontrolled cell losses. Far-red fluorescence makes DRAQ5<sup>™</sup> spectrally compatible with most visible range chromophores for multi-colour analysis. It is excited by blue through red laser lines, found on most flow cytometers. For convenience DRAQ5<sup>™</sup> is supplied in an aqueous ready-to-use formulation.

## **DRAQ5™** Product Features:

- far-red fluorescing live-cell permeant dsDNA probe
- water-soluble; easy-to-use straight from the fridge
- rapid, stoichiometric labelling of all nucleated cells
- compatible with BV / BUV, FITC & R-PE dyes
- excited by red or blue laser lines



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